



## Research Article

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# Effects of core iron oxide nanoparticles on microbial control and bacteriostatic activity against *Escherichia coli*, *Staphylococcus aureus* and *Mycobacterium smegmatis*

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## Abstract

The anti-bacterial activity of the Fe<sub>3</sub>/citric acid/cephalosporin nanoparticles (core) on *Staphylococcus aureus* (MTCC737), *Mycobacterium smegmatis* (MTCC994) and *Escherichia coli* (MTCC443) was tested using this system as an active compound carrier. We have describe the simple method to obtain iron nanoparticles with uniform size distribution by utilizing citric acid and as surface-capping agents the dimensions of Fe nanoparticles were in the 5-20 nm range and they were characterized by High Resolution Transmission Electron Microscopy (HRTEM).The particle size was tailored by tuning the concentration ratio of iron ions to carboxylic acid groups. Fe nanoparticles assisted by citric acid to analyze the interaction different type of cephalosporin against the these bacteria The antibacterial activity was observed in both, citric acid core nanoparticles/cephalosporin and core nanoparticles alone against these pathogen.

**Keywords:** Nanofluid, Magnetic, Fe nanoparticles, Antibacterial, Cephalosporin, HRTEM.

## Introduction

Magnetic nanoparticles have drawn much scientific interest by using magnetic nanoparticles for biological and medical purposes for a variety of studies.<sup>1</sup> Magnetic iron oxide based inorganic nanostructures materials have been synthesized and tested for various applications in medicine: as imaging agents, as heat mediators in hyperthermia treatments, in tissue repair, immunoassay, detoxification of biological fluids, cell separation, as magnetic guidance in drug delivery. The advantages of using these materials come from their magnetic properties, high surface area that provides higher sensitivity, better targeting and improvement of the colloidal stability of the nanostructures.<sup>2</sup> Important properties of the nanoparticles required for biomedical applications<sup>3, 4</sup> which are derived from a precise control of particle size, shape, dispersion and conditions that affect these properties. One area that is particularly promising is the use of magnetic nanoparticle systems for probing and manipulating biological systems.<sup>5</sup> Coating nanoparticles with natural or synthetic polymers or surfactants is a method that provides stability of the ferrofluid colloidal suspensions. Use of surfactants such as: citric acid, oleic acid, hexaldehyde or sodium carboxymethyl cellulose leads to highly dispersed and high quality nanoparticles with good biocompatibility and smaller particle size.<sup>4</sup>

Coated nanoparticles are important for their lower toxicity due to the presence of the biocompatible coating, and also due to the lower adsorption sites for proteins, ions and other components in medium.<sup>6</sup> Usually iron oxides Fe<sub>3</sub>O<sub>4</sub> or  $\gamma$ -Fe<sub>3</sub>O<sub>4</sub> are synthesized through the coprecipitation of Fe<sup>2+</sup> and Fe<sup>3+</sup> aqueous salt solutions<sup>5</sup>, by addition of a base. Properties of nanoparticles such as size, shape and composition, are influenced by the type of salt, pH, ions ratio and ionic strength of the medium.<sup>7</sup> Some methods use magnetotactic bacteria (MTB)<sup>8</sup> that are able to internalize Fe and convert it into magnetic nanoparticles, in the form of either

magnetite ( $\text{Fe}_3\text{O}_4$ ) or greigite ( $\text{Fe}_3\text{S}_4$ )<sup>9</sup>, Electrochemical preparation has also been used in situ for synthesis of core-shelled  $\text{Fe}_3\text{O}_4$  nanoparticles.<sup>10</sup> Sun *et al.*<sup>11, 12</sup> developed a thermal decomposition method that uses a Fe/acac salt, 1, 2-hexadecanediol, oleic acid, oleylamine, and biphenyl ether mixture to obtain nanoparticles that are further used for silver coating in order to improve bacterial activity and paramagnetic properties of the nanostructures.<sup>13</sup> A system that uses a combination of magnetron sputtering and gas-aggregation techniques produces Fe nanoclusters of variable controlled mean size (diameters from 2 to 100nm) and high magnetic moments for biomedical applications.<sup>14</sup> Magnetic temperature sensitive liposomes based on  $\text{Fe}_3\text{O}_4$  could be used to generate heat to achieve a certain temperature and during this time encapsulated anticancer drugs are released.<sup>15</sup>

In this study, new magnetic core-shell iron oxide base nonmaterial was obtained, adapting the coprecipitated method in order to improve colloidal dispersion and to control particles size.<sup>16</sup> Small size of nanoparticles was also on purpose, due to the possibility of targeting through blood barriers.<sup>17</sup> Citric acid was used as surfactant for coating the  $\text{Fe}_3\text{O}_4$  nanoparticles, followed by adsorption-coating with four different cephalosporins. The bacterial activity was tested on three different bacteria: *E coli*, *S. aureus* and *M. smegmatis*

## Materials and Methods

### Materials

*Staphylococcus aureus* (MTCC737), *Mycobacterium smegmatis* (MTCC994) and *Escherichia coli* (MTCC443) has been brought from IMTECH Chandigarh, India

### Iron (Fe) nanoparticles synthesis

Fe nanoparticles were synthesized at room temperature from  $\text{FeCl}_3$  using  $\text{NaBH}_4$  as a reducing agent and  $\text{PdCl}_2$  which serves as a nucleating agent. The concentration ratio ( $R+/-$ ) of iron ions ( $\text{Fe}^{3+}$ ) to citric acid monohydrate (CA) was systematically varied and its effect on the resulting nanoparticles size was studied. The concentration ratio ( $R+/-$ ) is defined as the ratio of total positive ( $\text{Fe}^{3+}$ ) to negative charges (CA and OA) which can be present in solution. The  $R+/-$  is calculated to be  $3[\text{Fe}^{3+}]/([\text{OA}] + 3[\text{CA}])$ . In a typical nanoparticles synthesis procedure ( $R+/- = 0.86$ ), 0.16 mmol  $\text{FeCl}_3$ , 0.16 mmol CA, 0.08 mmol OA and 0.1 ml  $\text{PdCl}_2$  solution (0.01 M) were dissolved in a mixed solvent (150 ml water and 40 ml ethanol). After purging with a gas mixture [95% argon (Ar) and 5% hydrogen ( $\text{H}_2$ )] for 30 min, 0.06 g  $\text{NaBH}_4$  of 10 ml water solution was added in one shot and after that mixture immediately became black, indicating the formation of Fe nanoparticles.

### Characterization of Iron oxide

Transmission Electron Microscopy (TEM) images were taken on a Phillips-CM200 electron microscope at 200 kV. One drop of

suspended samples (solvent-ethanol) was deposited on carbon coated copper grid and dried. TEM was used to observe the morphology of the nanoparticles. The particle size is reported as the average observed size ( $D_{avg}$ ), which is the average particle size of approximately 1000 individual particles from multiple TEM images.<sup>18, 19</sup>

### Determination of anti-bacterial activity

The qualitative antibiogram interpretation process was done according to international standard.<sup>20</sup> Inoculum preparation has been performed by making suspensions from 2-3 colonies isolated in physiological serum; the suspension turbidity has been done either nephelometric controlled or by comparison with standard tubes. Seeding was done by proper medium was chosen according to the tested bacterial species. Incubation has been depending upon the bacterial species: in normal atmosphere, 37°C, 20-24 hours for *E.coli*, *M. smegmatis* and *S.aureus*; *Interpretation*: a confluent bacterial culture appeared; inhibition zones appeared around the micro pills (the lack of the bacterial growth); the diameters of the inhibition zones were read, taking into account the used antibiotic, the quantity of the antibiotic in the pill and the tested bacterial species. The sensitivity diameters were compared with the standard ones.

## Results and Discussion

### Fe nanoparticles synthesis

Citric acid [CA] has been traditionally utilized as surface capping agent to control the formation of metallic nanoparticles such as cobalt<sup>18</sup>, gold<sup>21</sup>, palladium<sup>22</sup>, and silver<sup>23</sup>. In our Fe nanoparticles synthesis we also utilized  $\text{PdCl}_2$ , Which serves as a nucleating agent, along with CA to control Fe nanoparticles formation.<sup>24</sup> Figure 1(a) shows that the Fe particle size ranges from 200 to 20 nm as the concentration of  $\text{Fe}^{3+}$  to CA remained at 10. This result revealed that the amount of CA cannot control Fe nanoparticles formation when there is more  $\text{Fe}^{3+}$  than CA present in the solution. When the amount of CA was increased and  $R+/-$  became 0.7, uniform Fe nanoparticles with an average size of 11 nm were successfully synthesized (figure 1(b)). These results suggest that a greater amount of CA (or lower  $R+/-$ ) leads to smaller particles with more uniform size distribution, which is consistent with other reports.<sup>19, 22, 23</sup>

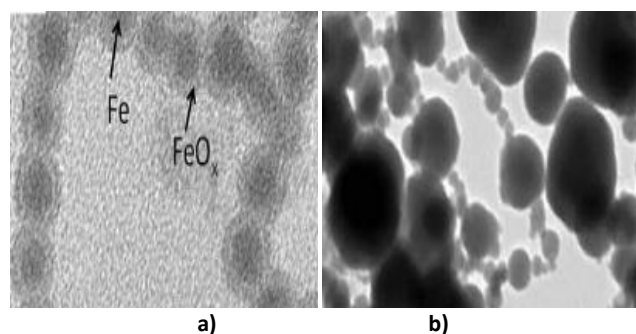


Figure 1: TEM images of Fe nanoparticles synthesized at formulation ratios of  $\text{Fe}^{3+}$  to citric acid

### Determination of anti-bacterial activity

Bacteriostatic activity by cephalosporin drug alone (table 1) and with nanoparticles core alone (table 2) were tested with three different bacteria, *E. coli*, *M. smegmatis* and *B. subtilis*. It was observed that, for the same time interval, the inhibition zone

diameters for cephalosporins conjugated with nanoparticles (Table 3) were higher from both the cephalosporin and nanoparticles core alone. This leads that the nanofluid increase potency of drug as it act as carrier and itself act as bacteriostatic activity.

**Table 1:** Inhibition zone diameter on *E. coli*, *Mycobacterium smegmatis* and *S. aureus*

Cephalosporins	Inhibition zone diameter [mm] on <i>Escherichia coli</i>	Inhibition zone diameter [mm] on <i>Staphylococcus aureus</i>	Inhibition zone diameter [mm] on <i>Mycobacterium smegmatis</i>
<i>Cefoperazone</i>	21	22	17
<i>Cefotaxime</i>	26	28	16
<i>Ceftriaxone</i>	28	27	19
<i>Cephachlor</i>	19	29	16

**Table 2:** Inhibition zone diameter on *E. coli* and *S. Aureus* on Fe<sub>3</sub>O<sub>4</sub> /citric acid (Core)

Nanofluid	Inhibition zone diameter [mm] on <i>Escherichia coli</i>	Inhibition zone diameter [mm] on <i>Staphylococcus aureus</i>	Inhibition zone diameter [mm] on <i>Mycobacterium smegmatis</i>
Fe <sub>3</sub> O <sub>4</sub> -citric acid core/shell	15	12	10

**Table 3:** Inhibition zone diameters of cephalosporins extra-shelled Fe<sub>3</sub>O<sub>4</sub>

Cephalosporins adsorption-shelled Fe <sub>3</sub> O <sub>4</sub>	Inhibition zone diameter [mm] on <i>Escherichia coli</i>	Inhibition zone diameter [mm] on <i>Staphylococcus aureus</i>	Inhibition zone diameter [mm] on <i>Mycobacterium smegmatis</i>
<i>Cefoperazone</i>	24	27	15
<i>Cefotaxime</i>	28	26	13
<i>Ceftriaxone</i>	31	24	16
<i>Cephachlor</i>	21	22	11

### Conclusions

We have demonstrated the feasibility of preparing low loss, Fe nanoparticles polymer composites. We have design a simple method to obtain iron nanoparticles with uniform size distribution by utilizing citric acid and as surface-capping agents. The particle size was fabricated by tuning the concentration ratio of iron ions to carboxylic acid groups. The Fe nanoparticles were successfully coated by a layer of silica to prevent Fe cores from oxidizing. Adsorption process is a convenient way to obtain a different type cephalosporin drug carrier's nanoparticles core. These core nanoparticles drug carrier not only useful in bacteriostatic effect against *E. coli*, *M. smegmatis* and *S. Aries* but it also enhances the efficacy and potency against

cephalosporin antibiotic. The small size of the core nanoparticles makes possible the delivery of the antibiotic when targeting certain organs like the brain, liver and kidney. The method of drug encapsulation with core nanoparticles will be very useful for drug delivery system against visceral leishmania<sup>25</sup> and biomedical research. It also applicable where the potency of drug is very low that can not able to elicit the enough response to inhibit the growth of pathogen.

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## References

1. C. P. Bean and J. D. Livingstone. Superparamagnetism. *J. Appl. Phys.* 1959; 30: 120S.
2. P. Wust, U. Gneveckow, M. Johannsen, D. Böhmer, T. Henkel, F. Kahmann, *et al.*, Magnetic nanoparticles for interstitial hyperthermia—feasibility, tolerance and achieved temperatures. *Int J Hyperthermia.* 2006 Dec;22(8):673-685.
3. S. Müller, Magnetic fluid hyperthermia therapy for malignant brain tumors—an ethical discussion. *Nanomedicine: Nanotechnology, Biology, and Medicine* 2009; 5: 387-393.
4. Byrappa K, Ohara S, Adschiri T. Nanoparticles synthesis using supercritical fluid technology- towards biomedical applications. *Advanced Drug Delivery Reviews.* 2008;60:299-327.
5. K. Raj, B. Moskowitz, R. Casciari, Advances in ferrofluid technology. *J. Magn. Mater.* 1995; 149: 174-180.
6. M. Mahmoudi, A. Simchi, M. Imani, M.A. Shokrgozar, A.S. Milani, U. O. Häfeli, P. Stroeve, A new approach for the *in vitro* identification of the cytotoxicity of superparamagnetic iron oxide nanoparticles. *Colloids and Surfaces B: Biointerfaces* 2010; 75: 300-309.
7. A. K. Gupta, M. Gupta, Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. *Biomaterials* 2005; 26(18): 3995-4021.
8. Mandal D, Bolander ME, Mukhopadhyay D, Sarkar G, Mukherjee P. The use of microorganisms for the formation of metal nanoparticles and their application. *Applied Microbiology and Biotechnology* 2006; 69: 485-492.
9. Xie, J. ; Chen, K. ; Chen, X. Production, Modification and Bio-Applications of Magnetic Nanoparticles Generated by Magnetotactic Bacteria. *Nano research* 2009; 2: 261-278.
10. J. Chumming, L. Xiangqin, Electrochemical synthesis of Fe<sub>3</sub>O<sub>4</sub>-PB nanoparticles with core-shell structure and its electrocatalytic reduction toward H<sub>2</sub>O<sub>2</sub>. *Solid State Electrochem.* 2009; 13: 1273-1278.
11. S. H. Sun, H. Zeng, Size-controlled synthesis of magnetite nanoparticles. *J. Am. Chem. Soc.* 2002; 124: 8204.
12. Sun S1, Zeng H, Robinson DB, Raoux S, Rice PM, Wang SX, Li G. Monodisperse MFe<sub>2</sub>O<sub>4</sub> (M = Fe, Co, Mn) nanoparticles. *J Am Chem Soc.* 2004 Jan 14;126(1):273-279..
13. B Chudasama, AK Vala, N Andhariya, RV Upadhyay, RV Mehta. Enhanced antibacterial activity of bifunctional Fe<sub>3</sub>O<sub>4</sub>-Ag core-shell nanostructures. *Nano Research* 2009; 2(12): 955-965.
14. Y Qiang, J Antony, A Sharma, J Nutting, D Sikes, D Meyer. Iron/iron oxide core-shell nanoclusters for biomedical applications. *Journal of Nanoparticle Research* 2006; 8(3-4): 489-496.
15. M. Babincova, D. Kaljarova, G. Millagros Castilla Bautista, P. Babinec, Contactless radiocontrol of Colchicine and Cisplatin release from magneto-liposomes: new technologies can improve the performance of old drugs. *Digest Journal of Nanomaterials and Biostructures* 2009; 4(3): 395-401.
16. A. Goodarzi, Y. Sahoo, M.T. Swihart, P.N. Prasad; Aqueous Ferrofluid of Citric Acid Coated Magnetite Particles. *Materials Research Society, Symposium Proceedings* 2004; 789: 129-134.
17. O. Veiseh, J. W. Gunn, M. Zhang, Design and fabrication of magnetic nanoparticles for targeted drug delivery and imaging. *Advanced drug delivery reviews* 2006; 62(3): 284-304.
18. Kobayashi Y, Horie M, Konno M, Rodriguez-González B, Liz-Marzán LM: Preparation and properties of silica-coated cobalt nanoparticles. *J Phys Chem B* 2003; 107: 7420.
19. Z. Zhang, F. Zhou, E. J. Laverna, "On the analysis of grain size in bulk nanocrystalline materials via X-ray. *Metall Mater Trans.* 2003; A, 34A: 1349-1355.
20. <http://www.clsj.org/>
21. Enustun B V and Turkevich J. Coagulation of colloidal gold. *J. Am. Chem. Soc.* 1963; 85: 3317-3328.
22. Turkevich J and Kim. Palladium-preparation and catalytic properties of particles of uniform size. *Science* 1970; 169: 873.
23. Henglein A and Giersig M. Formation of colloidal silver nanoparticles. Capping action of citrate, *J. Phys. Chem. B* 1999; 103: 9533-9539.
24. P Toneguzzo, G Viau, O Acher, F Fiévet-Vincent, F Fiévet. Monodisperse ferromagnetic particles for microwave applications. *Advanced Materials* 1998; 10(13): 1032-1035.
25. Rishikesh K, Ganesh GC, Krishna P, VNR Das and Pradeep D. Targeting Leishmaniasis: Review Multifunctional Aspect of Nanoconjugates and Nanoparticles. *IJIRS* 2013; 2(12): 99-110.